

# **Macroscopic forces generated by cell-matrix interactions**

- I. Cells generate forces after becoming attached to a matrix.**
- II. How do cells attach to a matrix?**
- III. Cell-matrix interactions control the spontaneous closure of wounds in organs.**
- IV. What happens when wound closure occurs by induced regeneration?**

# III. Cell-matrix interactions control the spontaneous closure of wounds in organs.

- Outline.
- What are the different types of tissue and organ injury? How does the organism respond to injury?
- The irreversibility of injury
- Mechanism of contraction and scar formation
- Quantitative measurement of healing modes: The defect closure rule

# **Description of response to injury: Contraction and scar formation following trauma**

## **A. Sources of trauma: energy sources.**

- **mechanical: deep cut, laceration, surgery**
- **thermal: fire, hot water.**
- **electromagnetic: UV, electrical discharge**
- **nuclear: radiation therapy.**

# **Contraction and scar formation following trauma (cont.)**

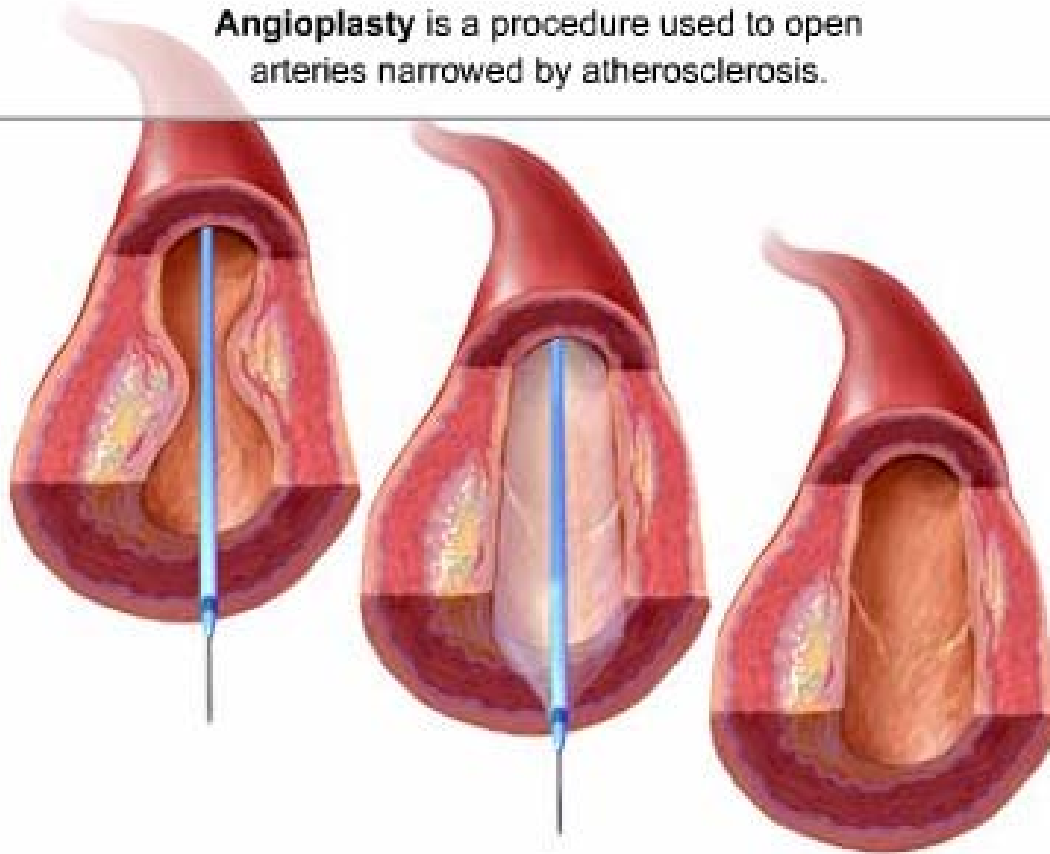
## **B. Outcome of trauma, scar formation and contraction**

- **on finger joint it prevents movement (“contracture”)**
- **in peripheral nerves (neuroma) it prevents conduction of electric signals (paralysis)**
- **in neck of face it creates serious problems of social acceptance**
- **around suture points (e.g., following caesarian section)**
- **surgical adhesions prevent normal function of intestines**

Image removed for copyright reasons.  
National Football League poster warning about the  
dangers of tackling with the helmet.

**Angioplasty** is a procedure used to open arteries narrowed by atherosclerosis.

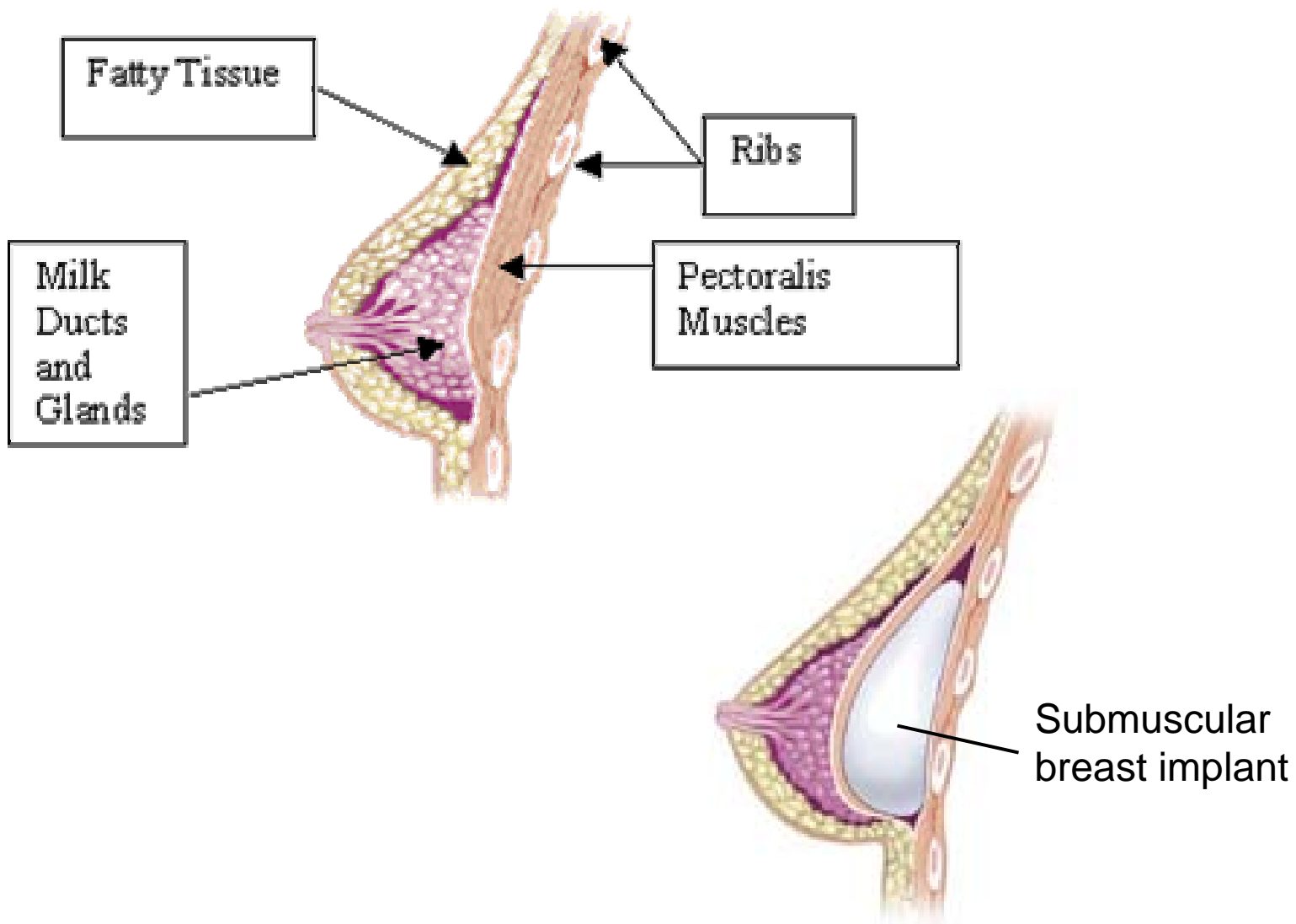
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Source: U.S. FDA. "Cardiac Angioplasty Device."  
<http://www.fda.gov/hearthealth/treatments/medicaldevices/angioplasty.html>

## **C. Scar formation around “inert” implant**

- 1. Scarring following implantation of any nondegradable prosthesis (e.g., silicone, polyethylene)**
- 2. Constrictive scar tissue (fibrous capsule) around implant causes chronic pain and implant deformation**
- 3. Implantation of nonporous, biodegradable sheet under skin (subcutaneous) leads to encapsulation of implant by fibrotic tissue. No capsule formation around identical implant, except for being porous.**
- 4. Implants are often supported mechanically by contraction and scar formation around them.**



Source: U.S. FDA. "Making an Informed Decision: Saline-Filled Breast Implant Surgery – 2004 update."  
[http://www.fda.gov/cdrh/breastimplants/labeling/inamed\\_patient\\_labeling\\_5900.html](http://www.fda.gov/cdrh/breastimplants/labeling/inamed_patient_labeling_5900.html)



## **D. Chronic scar formation**

- **Scar formation and contraction result from chronic trauma; or from acute or chronic inflammation caused by various agents**
- **Scar takes different names depending on medical specialty**
- **Examples: 1. Scarred heart valve due to incidence of rheumatic fever leads, e.g., to valve stenosis or to leakage.**  
**2. Necrosis (death) of myocardium (infarct) due to interruption of oxygen supply (clogged arteries) interferes with electrical conduction of heart muscle**  
**3. Obstruction of intestinal tract, due to chronic inflammation, leads to digestive problems (e.g., duodenal ulcer with gastric outlet obstruction)**  
**4. Fibrotic liver (cirrhosis) prevents liver function**

**scarred heart muscle  
(heart attack)**

**scarred liver  
(cirrhosis)**

**scarred kidney  
(infection)**

Diagram removed due to copyright considerations.  
See Figure 1.3 in [Yannas 2001]: Yannas, I. V. *Tissue and Organ Regeneration in Adults*. New York: Springer, 2001.

**scarred cornea  
(infection)**

**scarred heart valve  
(rheumatic fever)**

# **The irreversibility of injury**

# Amphibian: reversible injury

**Spontaneous  
regeneration  
of amputated  
limb in the  
newt (a small  
amphibian)  
occurs  
independently  
of severity of  
injury**

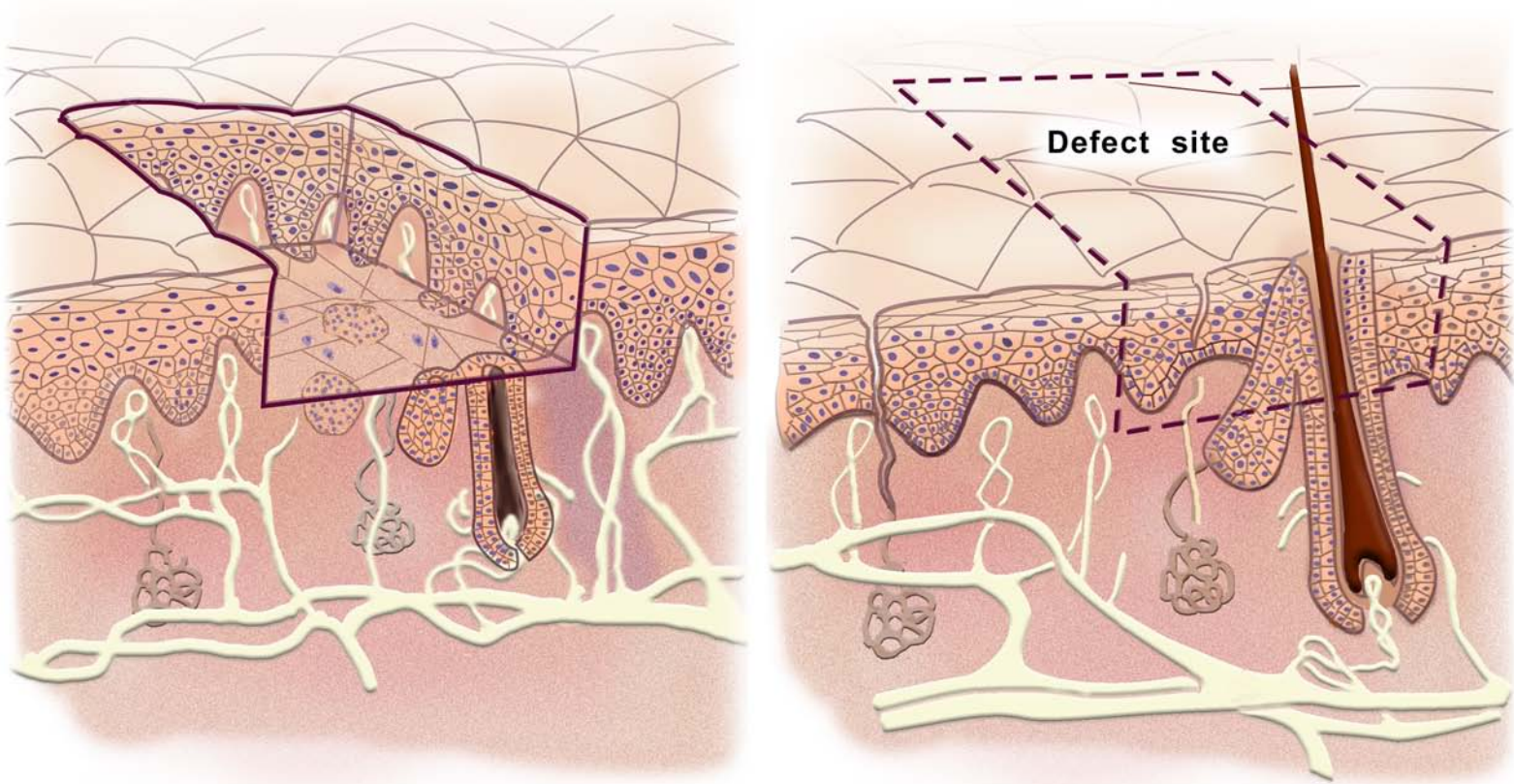
Diagram removed for copyright reasons.  
See Figure 1.1 in [Yannas 2001].

# Liver: Reversible or irreversible injury?

**The healed liver has the same mass, but a different shape (resected lobes are not regenerated), than the intact organ**

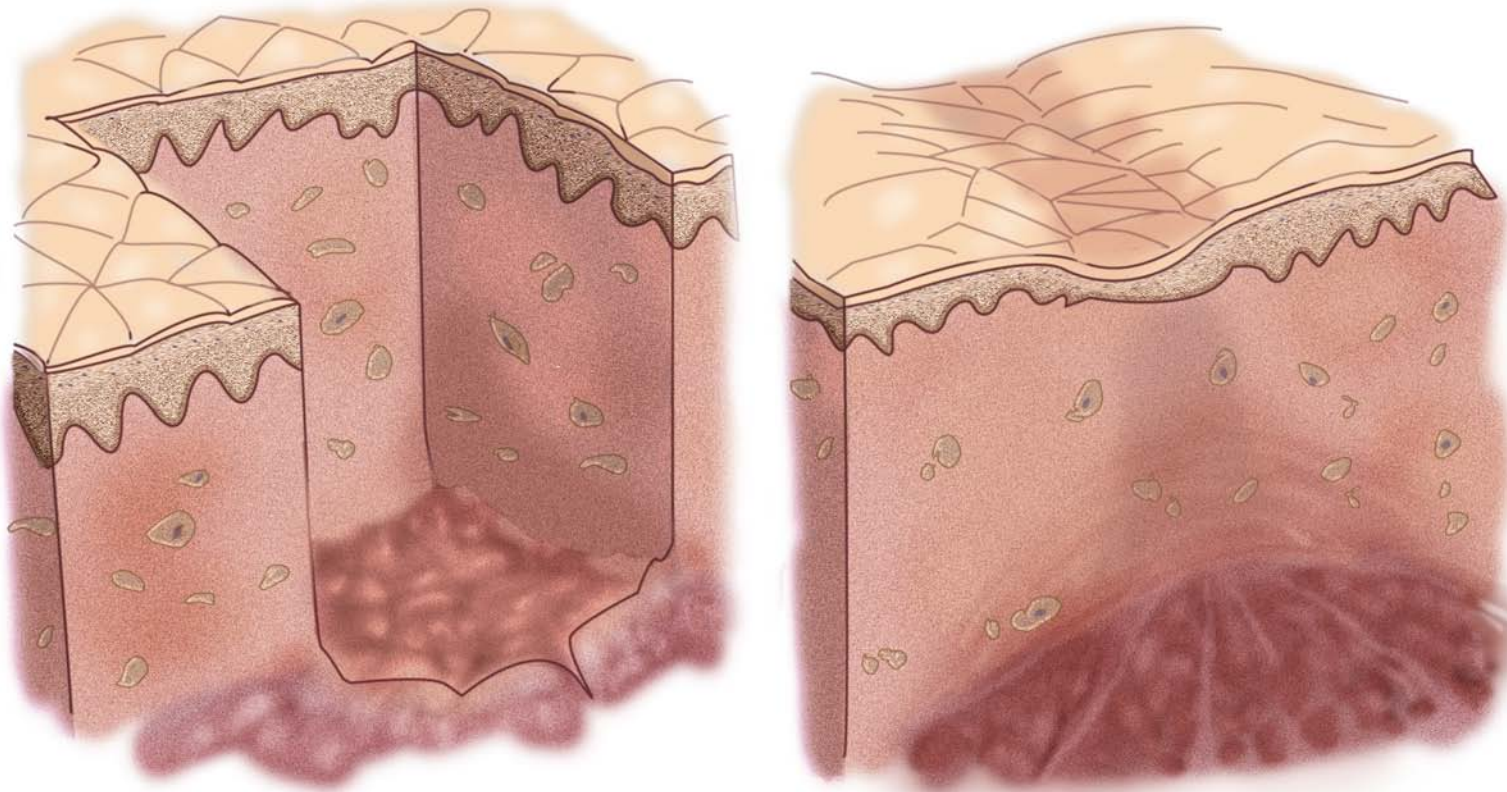
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See Figure 1.2 in [Yannas 2001].

# Skin: reversible injury



**Spontaneous regeneration of excised epidermis**

# Skin: irreversible injury



**Spontaneous healing of skin excised to full thickness by contraction and scar formation. The dermis does not regenerate.**

# Peripheral nerve: reversible injury

Mildly crushed  
nerve heals  
spontaneously  
by  
regeneration

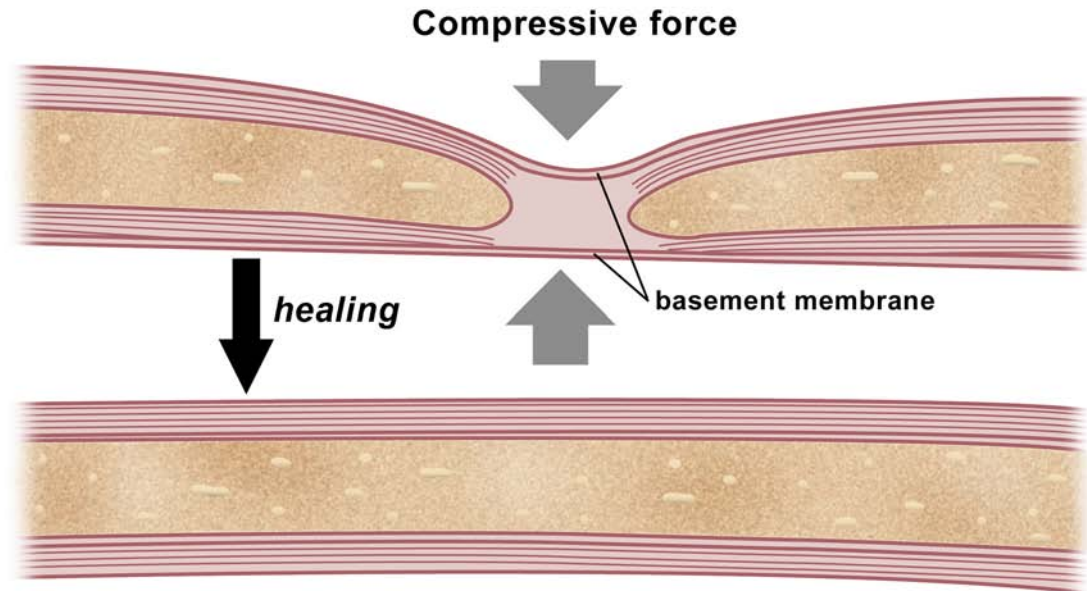
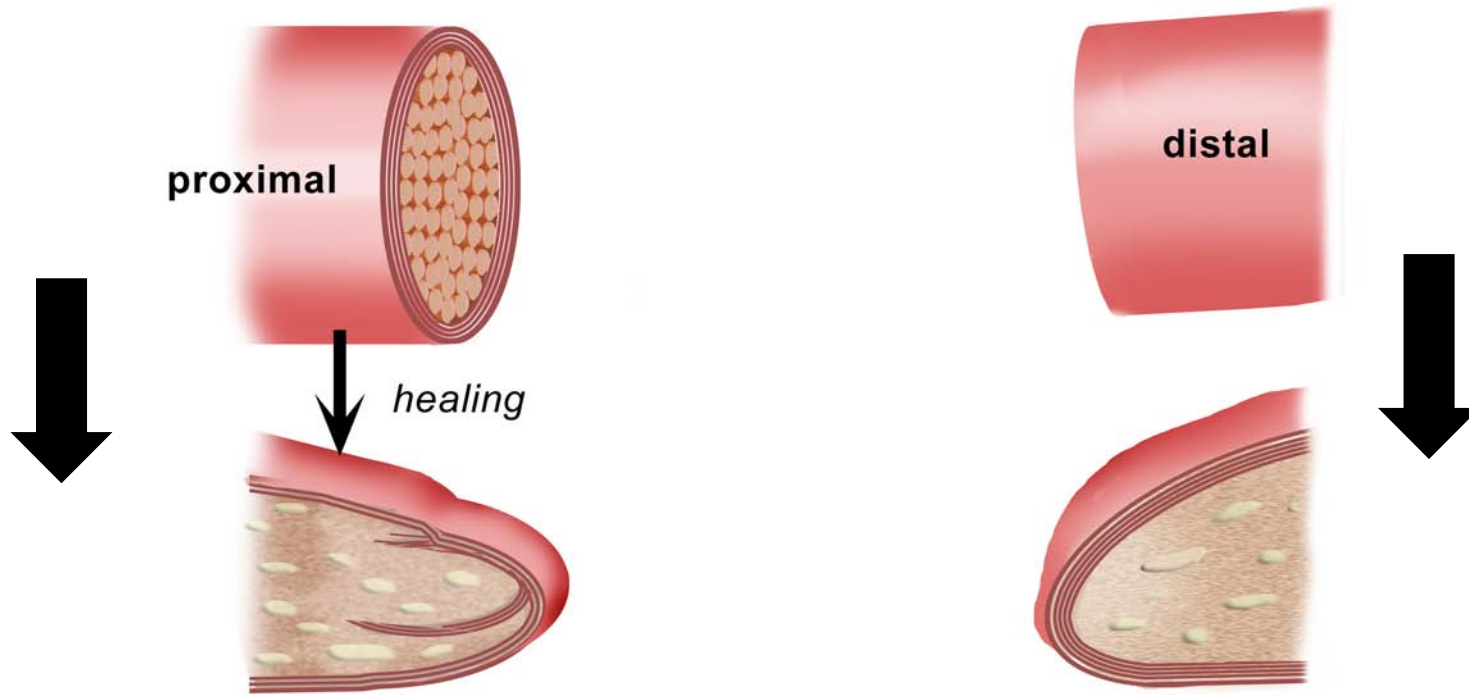


Figure by MIT OCW. After Figure 2.3 in [Yannas 2001].

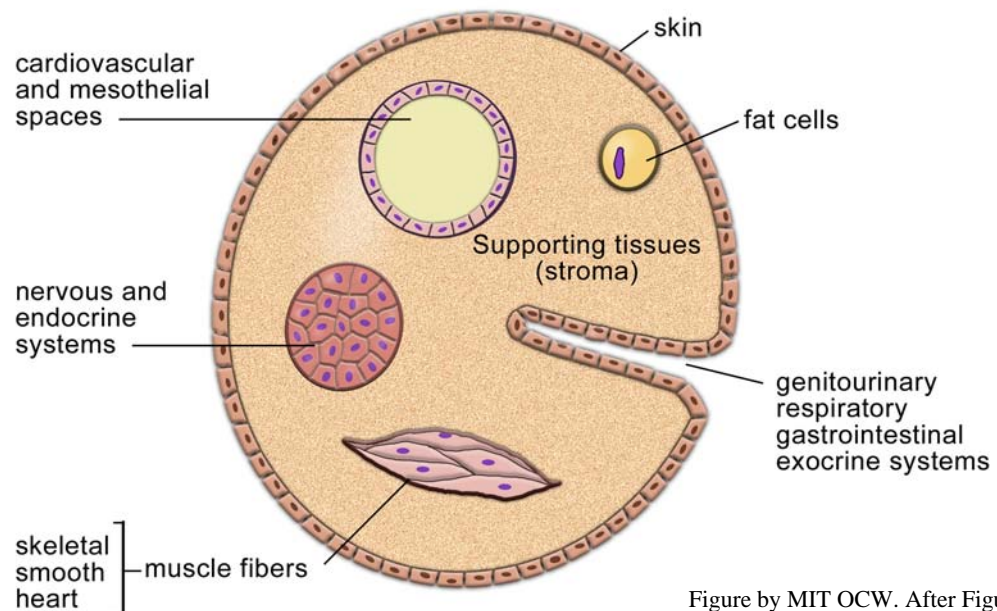
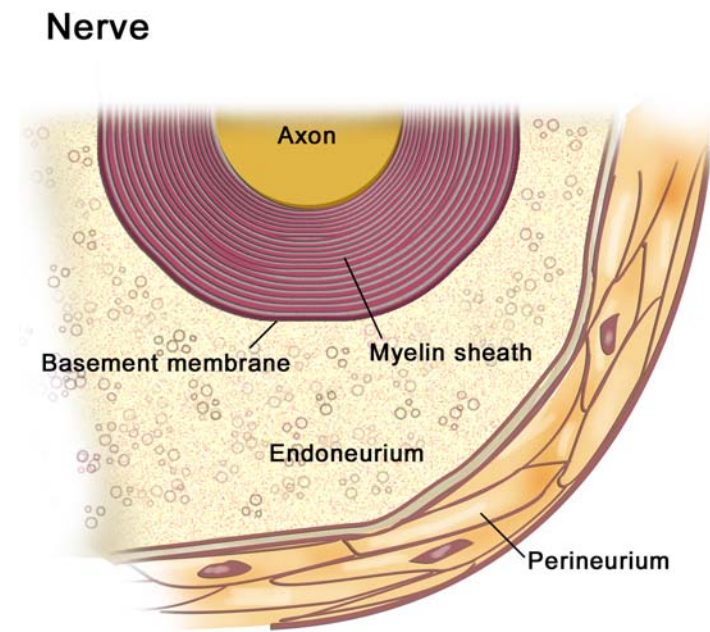
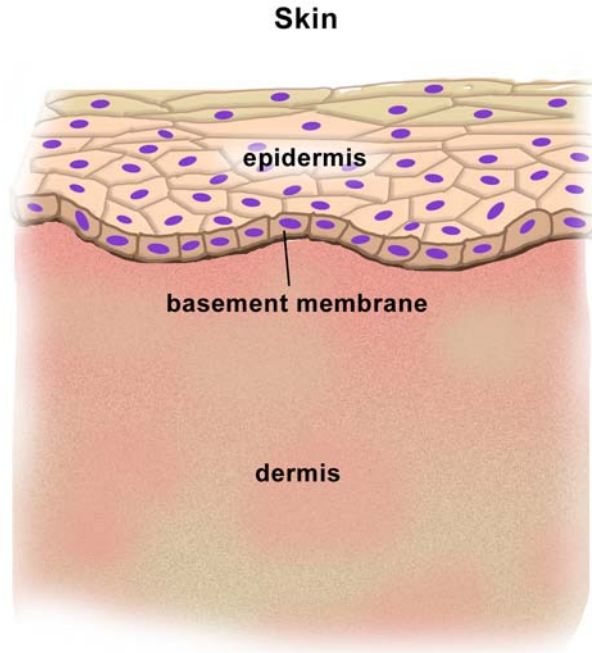


# Peripheral nerve: irreversible injury



**Transected nerve heals spontaneously by contraction and neuroma (neural scar) formation. No reconnection of stumps.**

# Generalized view of reversibility of injury based on rule of tissue triad



**Cartoon of “organism” shows that basement membrane (thick solid line) appears in almost all organs**

Figure by MIT OCW. After Figure 2.7 in [Yannas 2001].

**Healing rules: epithelial tissue and BM, spontaneously regenerative; stroma, nonregenerative**

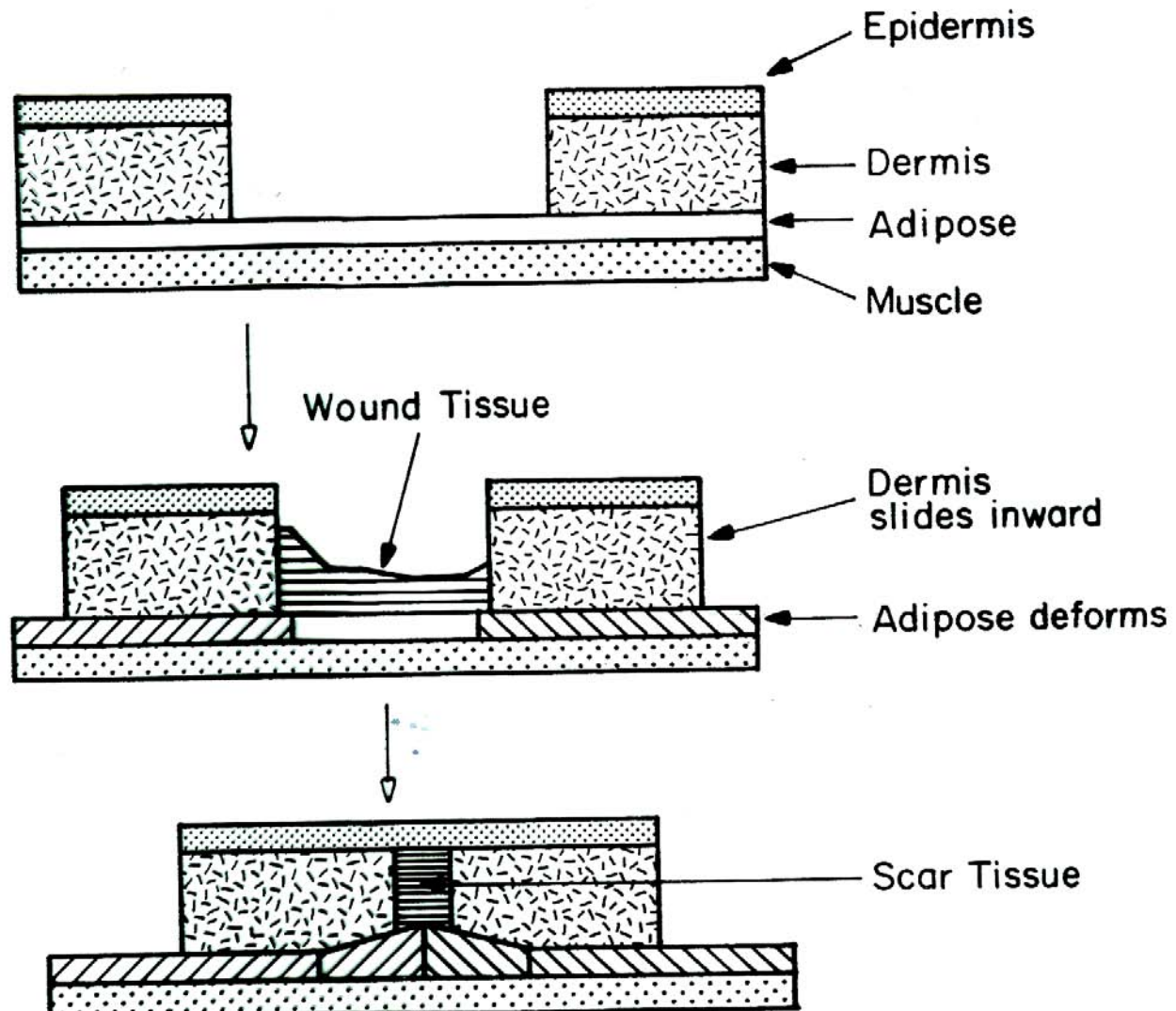
	<b>Regenerative tissues. Reversible injury. No contraction.</b>	<b>Nonregenerative tissues. Irreversible injury. Contraction +scar.</b>
<b>SKIN</b>	epidermis	dermis
	BM	
<b>NERVE</b>	myelin	endoneurial stroma
	BM	
<b>GENERAL ORGAN</b>	epithelial tissue BM	stroma

# **Mechanism of contraction and scar formation**

## **Mechanism of contraction and scar formation**

- 1. Movement of tissue from periphery of wound toward center**
- 2. Contractile fibroblasts (myofibroblasts) may initiate contraction; they almost certainly propagate contraction**
- 3. Collagen fibers in scar are highly oriented in the plane of the wound.**
- 4. Collagen fibers synthesized by FB and extruded outside with fiber axis parallel to long cell axis. Fiber orientation is replica of cell axis orientation during scar synthesis.**
- 5. Collagen fiber orientation in scar is in the plane of the wound, suggesting the presence of a plane stress field during scar synthesis.**
- 6. Regeneration templates cancel out mechanical field, leading to fiber synthesis in random orientation.**

## Deformation of Perilesional Tissues During Contraction



Four diagrams of basal cells removed for  
copyright reasons.

From Asmussen and Sollner,  
*Lume* 29:3, 1993

# Myofibroblast detected with antibody to $\alpha$ -SM actin

Diagram removed for copyright reasons.

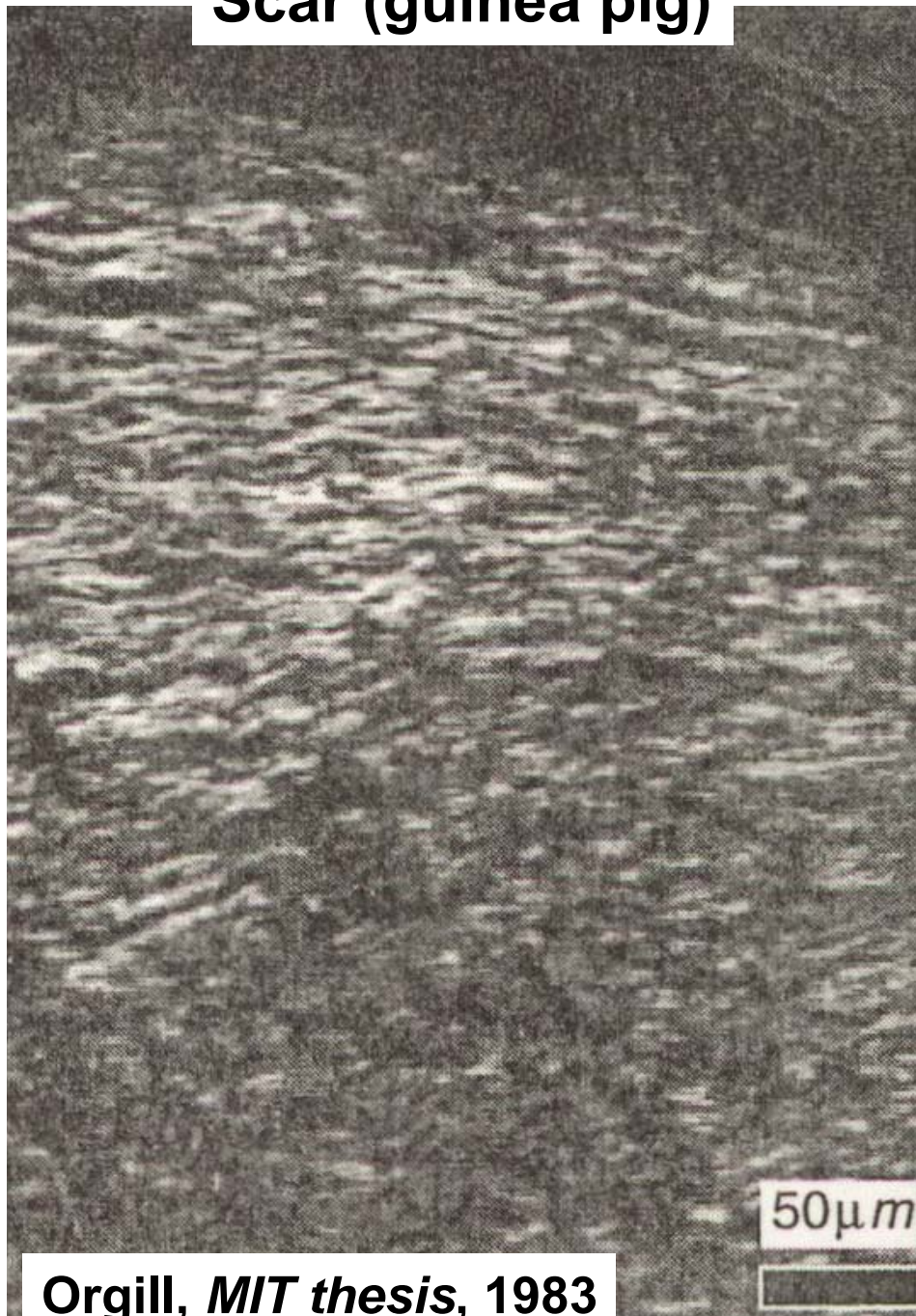


**Regenerated dermis  
(guinea pig)**



50  $\mu m$

**Scar (guinea pig)**



50  $\mu m$

**Orgill, *MIT thesis*, 1983**

## Measure S (quantitative assay)

Diagram & photo removed for copyright reasons.  
See Figure 4.7 in [Yannas 2001]

Original source: Ferdman and Yannas,  
*J. Invest. Dermatol.*, 1993

# Unit cell processes of scar formation

- Wound healing can be summed up as a sequence comprising an inflammatory response, fibroplasia, epithelialization, wound contraction and scar maturation. The following sequence of unit cell processes is a hypothetical and highly simplified model of certain aspects of wound healing in the dermal layer. Epithelialization and scar maturation are entirely omitted in the model below:
- $n \text{ Platelets} + \text{Quaternary-structured collagen} = [\text{Degranulation}] = \text{Thrombus} + \text{PDGF}^* \text{ (and TGF-}\beta^*)$

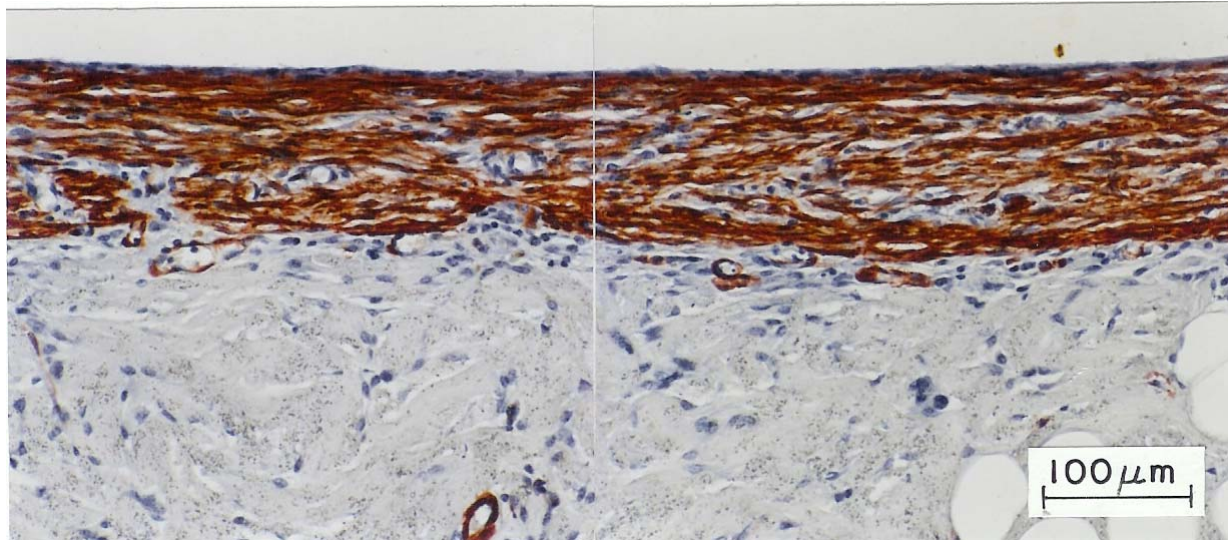
- 
- $\text{PDGF} + \text{Monocyte} = [\text{Differentiation}] = \text{Macrophage} + \text{PDGF}$
  - $\text{PDGF} + \text{Macrophage} + \text{ECM} = [\text{Collagenase}^{**} \text{ synthesis}] = \text{Solubilized ECM} + \text{Regulator}$
  - $\text{Regulator} + \text{Macrophage} + \text{Solubilized ECM} = [\text{Phagocytosis}] = \text{Degraded ECM} + \text{Regulator}$

## Unit cell processes of scar formation (Cont.)

- **PDGF + Fibroblast + ECM = [Mitosis] = Fibroblast proliferation + Regulator**
  - **Regulator + Fibroblast + ECM = [Synthesis] = Collagen I and III + Regulator**
- 
- **Composite unit cell process: Collagen synthesis + Angiogenesis = Granulation tissue**
- 
- **Regulator + Fibroblast + ECM = [Synthesis of  $\alpha$ -actin] = Contractile fibroblast ("Myofibroblast") + Regulator**
  - **Regulator + Myofibroblast + ECM = [Synthesis] = Scar tissue + Regulator**
  - **Regulator + Myofibroblast + ECM = [Contraction] = Closed wound + Regulator**
- 
- **Wound closes up. Myofibroblasts dedifferentiate to stable fibroblasts.**

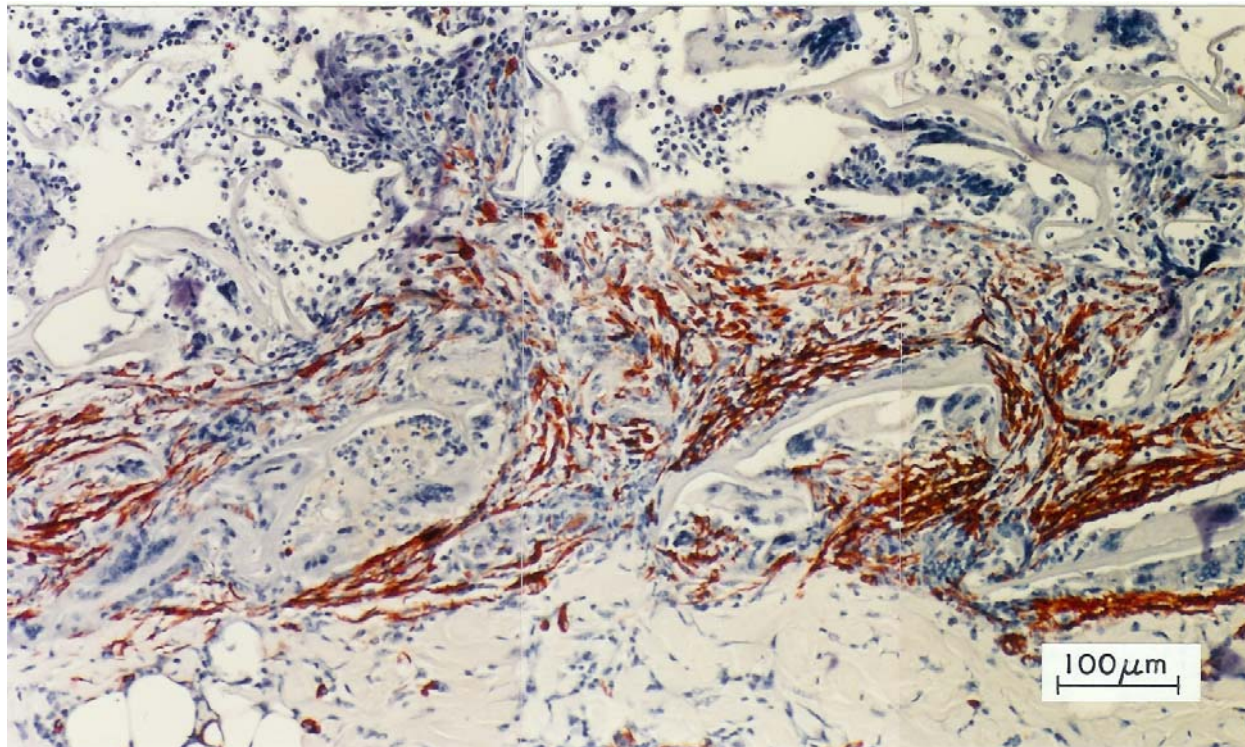
# Contraction blocked by scaffold (bottom)

**Ungrafted.  
Contracting  
vigorously.**



***Red-brown:  
stained with  
antibody to  
 $\alpha$ -SM actin.  
10 d***

**Grafted  
with DRT.  
No  
contraction.**



**From Troxel,  
*MIT thesis, 1994***

# **Quantitative measurement of healing response: The defect closure rule**

# Defect closure rule

Original wound area =  $A_o$

Wound area closed by contraction =  $A_c$

Wound area closed by scar formation =  $A_s$

Wound area closed by regeneration =  $A_r$

$$A_c + A_s + A_r = A_o \quad [1]$$

Dividing both sides of Equation [1] by  $A_o$  (normalization) we get the sum of the fractional areas:

$$A_c/A_o + A_s/A_o + A_r/A_o = 1 \quad [2]$$

Replacing the fractional areas with the symbols C, S and R, and multiplying by 100, we get the wound closure rule:

$$C + S + R = 100 \quad [3]$$

<b>Spontaneously healing defect</b>	<b>Configuration of final state</b>
general case	<b>[C, S, R]</b>
ideal fetal healing	<b>[0, 0, 100]</b>
dermis-free skin-- adult rodents	<b>[96, 4, 0]</b>
dermis-free skin-- adult human	<b>[37, 63, 0]</b>
peripheral nerve-- adult rat	<b>[96, 4, 0]</b>
conjunctiva-- adult rabbit	<b>[45, 55, 0]</b>

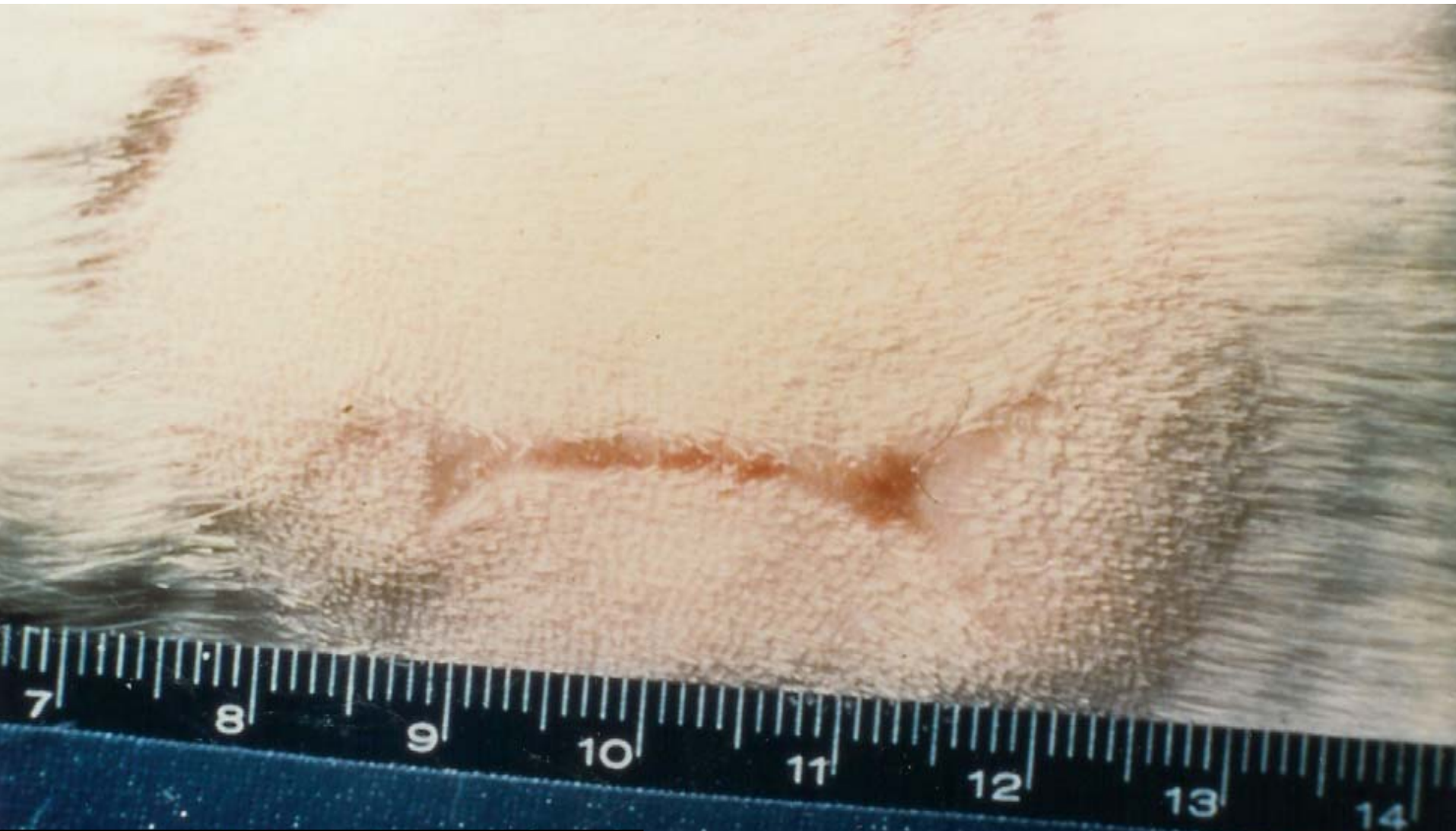


**Burn patient  
has closed  
severe skin  
wounds in  
neck partly  
by  
contraction  
and partly  
by scar**

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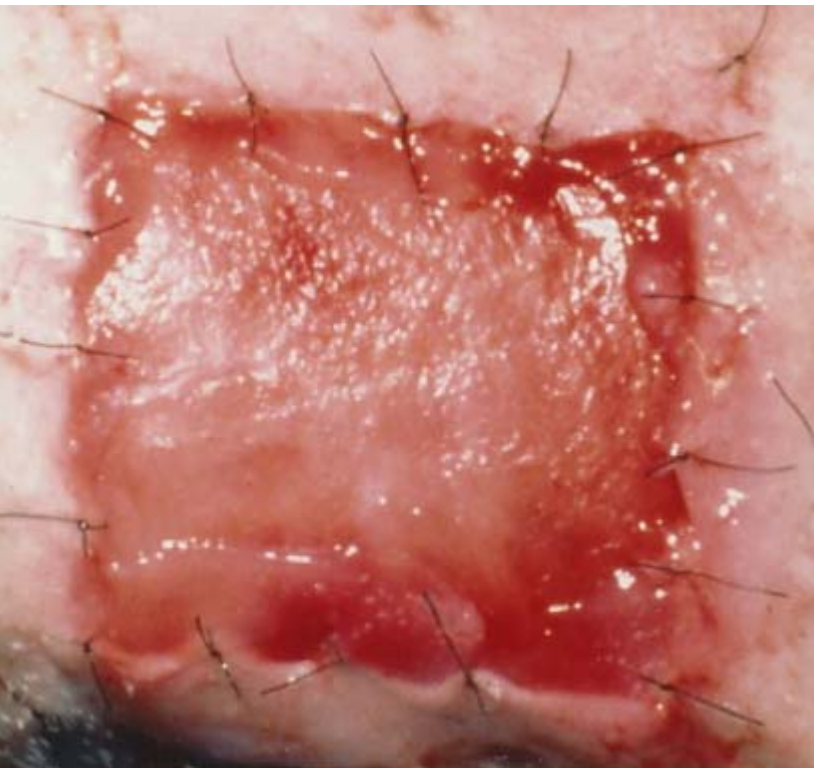
**Final state of healing of full-  
thickness skin wound in the  
human.**

Reference not  
recalled

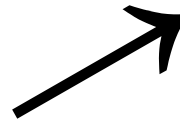


**Final state of healing of full-thickness skin wound in the guinea pig.**

**Orgill, *MIT thesis*, 1983**

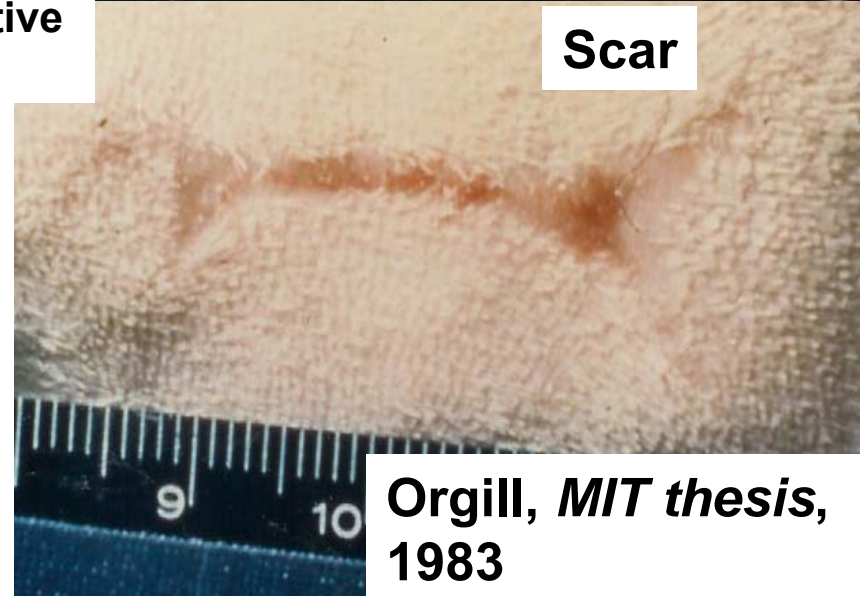
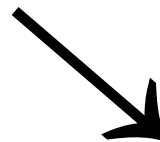


KC + DRT



Partly regenerated skin

KC + inactive scaffold



Scar

Full-thickness skin wound (guinea pig) grafted with keratinocytes (KC) and either dermis regeneration template (DRT) or inactive scaffold

Orgill, *MIT thesis*, 1983

# Measure C

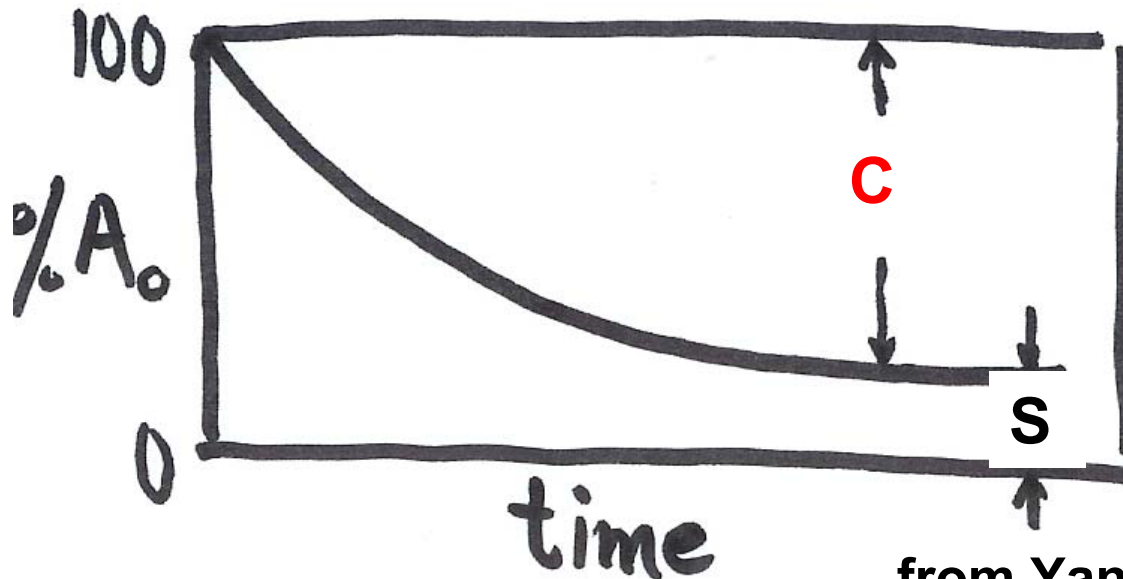
Graph removed for copyright reasons.  
See Figure 4.1 in [Yannas 2001]

# Kinetics of change in C

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See Figure 4.2 in [Yannas 2001]

Measurement of **C**, **S** and **R** in full-thickness skin wounds after wound has closed. Use only “final state” data!

Graph removed for copyright reasons.  
See Figure 4.3 in [Yannas 2001]



from Yannas' book, 2001

Figure removed for copyright reasons.  
See Table 4.1 in [Yannas 2001]